**NovoCyte Kl. II**

**V02**

**Relevant and important information for publication of flow cytometric data**

Remember to acknowledge the FACS Core Facility, Aarhus University in your publication and presentations (oral presentations and posters).

Send your paper to [*facs@au.dk*](mailto:facs@au.dk) for review and we will give you feedback within 2 workdays.

**Relevant information for methods and materials:**

*Instrument*

* NovoCyte 2100YB flow cytometer equipped with two lasers (488 nm and 561 nm) and 10 detectors (Agilent, Santa Clara, CA).

*Software*

* Software: NovoExpress (v. 1.6.2, Agilent, Santa Clara, CA).
* Other analysis software programs used: e.g. FlowJo or FCS Express (incl. version and company).

*Experimental*

* Antibodies (clone, isotype, manufacturer, fluorochrome) or fluorescent proteins used and in which filters they were detected.
* Antibody concentrations, staining conditions (incubation time, temperature, buffers, blocking etc.). State if antibody titration was performed.
* Controls included: E.g. compensation controls, biological controls, FMO controls, stimulation controls, mock controls.
* For compensation, state if you used beads or cells.
* Stop conditions: E.g. number of events in a specific gate or a sample volume.

Laser and filter overview on next page ⇒

**Abbreviations:**

|  |  |
| --- | --- |
| Forward Scatter | FSC |
| Side Scatter | SSC |

The table on the next page is an example of how to provide information about your experiment.

Enter the fluorochrome, fluorescent protein or DNA dye you used into the filter you used.

|  |  |  |  |
| --- | --- | --- | --- |
| Laser & power | Wavelength detected | Fluorochrome/  fluorescent protein | Antigen |
| 488 nm | 488/10 | FSC |  |
| 50 mw | 488/10 | SSC |  |
|  | 530/30 |  |  |
|  | 586/20 |  |  |
|  | 615/20 |  |  |
|  | 660/20 |  |  |
|  | 695/40 |  |  |
| 561 nm | 586/20 |  |  |
| 50 mw | 615/20 |  |  |
|  | 660/20 |  |  |
|  | 695/40 |  |  |
|  | 780/60 |  |  |