Penteon

V01

Relevant and important information for publication of flow cytometric data

Remember to acknowledge the FACS Core Facility, Aarhus University, in your publications and presentations (oral presentations and posters).
Send your papers to *facs@au.dk* for review and we will give you feedback within 2 workdays.

Methods and materials:

Instrument

- NovoCyte Penteon flow cytometer equipped with five lasers (349 nm, 405 nm, 488 nm, 561 nm and 637 nm) and 30 fluorescence detectors (Agilent, Santa Clara, CA).

Software

- Software: NovoExpress (v. 1.6.2, Agilent, Santa Clara, CA).
- Other analysis software programs used: e.g. FlowJo or FCS Express (incl. version and company).

Experimental

- Antibodies (clone, isotype, manufacturer, fluorochrome) or fluorescent proteins used and in which filters they were detected.
- Antibody concentrations, staining conditions (incubation time, temperature, buffers, blocking etc.). State if antibody titration was performed.
- Controls included: E.g. compensation controls, biological controls, FMO controls, stimulation controls, mock controls.
- For compensation, state if you used beads or cells.
- Stop conditions: E.g. number of events in a specific gate or a sample volume.

Laser and filter overview on next page ⇒

Abbreviations:

Forward Scatter	FSC
Side Scatter	SSC

The table below is an example of how to provide information about your experiment. Enter the fluorochrome, fluorescent protein or DNA dye you used into the filter you used.

Laser & power	Wavelength detected	Fluorochrome/ fluorescent protein	Antigen
349 nm	445/45		
20,2 mW	525/45		
	586/20		
	615/20		
	667/30		
	725/40		
	757-810		
405 nm	445/45		
100 mW	525/45		
	586/20		
	615/20		
	667/30		
	725/40		
	757-810		
488 nm	525/45	FSC	
100 mW	586/20		
	615/20		
	667/30		
	685-705		
	725/40		
561 nm	586/20	SSC	
100 mW	615/20		
	667/30		
	685-705		
	725/40		
	757-810		
637 nm	667/30		
100 mW	685-705		
	725/40		
	757-810		